

Type 100 Ti Rotor



Used In Beckman Coulter Class R and S Preparative Ultracentrifuges

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SAFETY NOTICE

This safety notice summarizes information basic to the safe use of the rotor described in this manual. The international symbol displayed above is a reminder to the user that all safety instructions should be read and understood before operation or maintenance of this equipment is attempted. When you see the symbol on other pages throughout this publication, pay special attention to the specific safety information presented. Observance of safety precautions will also help to avoid actions that could damage or adversely affect the performance of the rotor. This rotor was developed, manufactured, and tested for safety and reliability as part of a Beckman Coulter ultracentrifuge/rotor system. Its safety or reliability cannot be assured if used in a centrifuge not of Beckman Coulter's manufacture or in a Beckman Coulter ultracentrifuge that has been modified without Beckman Coulter's approval.



Handle body fluids with care because they can transmit disease. No known test offers complete assurance that such fluids are free of micro-organisms. Some of the most virulent—Hepatitis (B and C) viruses, HIV (I–V), atypical mycobacteria, and certain systemic fungi —further emphasize the need for aerosol protection. Handle other infectious samples according to good laboratory procedures and methods to prevent spread of disease. Because spills may generate aerosols, observe proper safety precautions for aerosol containment. Do not run toxic, pathogenic, or radioactive materials in this rotor without taking appropriate safety precautions. Biosafe containment should be used when Risk Group II materials (as identified in the World Health Organization *Laboratory Biosafety Manual*) are handled; materials of a higher group require more than one level of protection.



The rotor and accessories are not designed for use with materials capable of developing flammable or explosive vapors. Do not centrifuge such materials in nor handle or store them near the ultracentrifuge.



Although rotor components and accessories made by other manufacturers may fit in the Type 100 Ti rotor, their safety in this rotor cannot be ascertained by Beckman Coulter. Use of other manufacturer' components or accessories in the Type 100 Ti rotor may void the rotor warranty and should be prohibited by your laboratory safety officer. Only the components and accessories listed in this publication should be used in this rotor.



Place filled tubes in at least two opposing cavities. Make sure that filled containers are loaded symmetrically into the rotor and that opposing tubes are filled to the same level with liquid of the same density. Make sure that cavities in use have the proper floating spacers and/or spacers inserted (if applicable) before installing the rotor lid. Do not put spacers in cavities that do not contain tubes.



If disassembly reveals evidence of leakage, and pathogenic or radioactive materials are involved, you should assume that some fluid escaped the rotor. Apply appropriate decontamination procedures to the centrifuge and accessories.

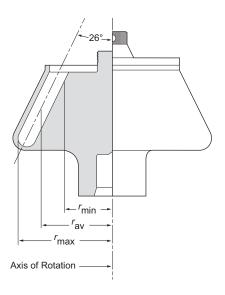


Never exceed the maximum rated speed of the rotor and labware in use. Refer to the section on RUN SPEEDS, and derate the run speed as appropriate.



Do not use sharp tools on the rotor that could cause scratches in the rotor surface. Corrosion begins in scratches and may open fissures in the rotor with continued use.

TYPE 100 Ti ROTOR



SPECIFICATIONS

Maximum speed
Density rating at maximum speed 1.2 g/mL
Relative Centrifugal Field* at maximum speed
At r_{max} (71.6 mm) 802 000 × g
At r_{av} (55.5 mm) 622 $000 \times g$
At r_{\min} (39.5 mm)
<i>k</i> factor at maximum speed
Conditions requiring speed reductions see RUN SPEEDS
Number of tube cavities
Available tubes see Table 1
Nominal tube dimensions (largest tube)
Nominal tube capacity (largest tube) 6.8 mL
Nominal rotor capacity
Approximate acceleration time to maximum speed
(fully loaded)
Approximate deceleration time from maximum speed
(fully loaded)
Weight of empty rotor
Weight of fully loaded rotor 4.1 kg (8.9 lb)
Rotor material titanium

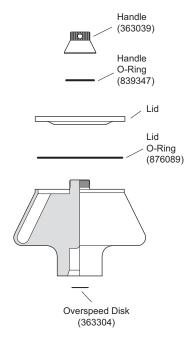
^{*} Relative Centrifugal Field (RCF) is the ratio of the centrifugal acceleration at a specified radius and speed $(r\omega^2)$ to the standard acceleration of gravity (g) according to the following formula:

$$RCF = \frac{r\omega^2}{g}$$

where r is the radius in millimeters, ω is the angular velocity in radians per second (2 π RPM /60), and g is the standard acceleration of gravity (9807 mm/s²). After substitution:

$$RCF = 1.12 r \left(\frac{RPM}{1000}\right)^2$$

DESCRIPTION



This rotor has been manufactured in a registered ISO 9001 or 13485 facility for use with the appropriately classified Beckman Coulter ultracentrifuges.

The Type 100 Ti, rated for 100 000 rpm, is a fixed angle rotor designed to centrifuge up to eight Quick-Seal® tubes at a 26-degree angle to the axis of rotation. Used in Beckman Coulter Class R and S preparative ultracentrifuges, the rotor develops centrifugal forces sufficient to pellet ribosomes or other micromolecules. It can also be used to purify and separate DNA, proteins, and other small particles in solution using density gradient techniques. Approximately 48 mL of gradient and sample can be centrifuged per run.

IIII NOTE

Certain models of the OptimaTM series ultracentrifuges may require software updates in order to run this rotor. Contact Beckman Coulter Field Service.

The rotor is made of titanium and is finished with black polyurethane paint. The lid and handle are made of anodized aluminum. O-rings made of Buna N rubber in the rotor handle and lid maintain atmospheric pressure inside the rotor during centrifugation, if they are properly lubricated.

Because of the weight of the rotor, drive pins are not required in the rotor drive hub cavity.

For overspeed protection, a photoelectric detector in Beckman Coulter ultracentrifuges monitors the overspeed disk on the rotor bottom and shuts down the run if speeds exceeding 100 000 rpm are detected.

See the Warranty at the back of this manual for warranty information.

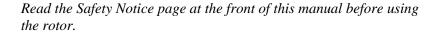
PREPARATION AND USE

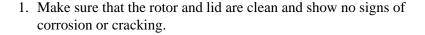
Specific information about the Type 100 Ti rotor is given here. Information common to this and other rotors is contained in the manual Rotors and Tubes for Preparative Ultracentrifuges (publication LR-IM), which should be used together with this manual for complete rotor and accessory operation. Publication LR-IM is included in the literature package shipped with this rotor manual.



Although rotor components and accessories made by other manufacturers may fit in the Type 100 Ti rotor, their safety in this rotor cannot be ascertained by Beckman Coulter. Use of other manufacturers' components or accessories in the Type 100 Ti rotor may void the rotor warranty and should be prohibited by your laboratory safety officer. Only the components and accessories listed in this publication should be used in this rotor.

PRERUN SAFETY CHECKS

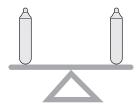




- 2. Make sure that the rotor is equipped with the correct overspeed disk (363304). If the disk is missing or damaged, replace it according to the instructions in *Rotors and Tubes*.
- 3. Check the chemical compatibilities of all materials used (refer to Appendix A in *Rotors and Tubes*).
- 4. Verify that only the tubes and accessories listed in Table 1 are being used.



ROTOR PREPARATION



For runs at other than room temperature, refrigerate or warm the rotor beforehand for fast equilibration.

- Be sure that metal threads in the rotor are clean and lightly but evenly lubricated with SpinkoteTM lubricant (306812). Also ensure that O-rings are lightly but evenly coated with silicone vacuum grease.
- 2. Load the filled and sealed tubes symmetrically into the rotor (see page 8 for tube information). If fewer than eight tubes are being

run, they must be arranged symmetrically in the rotor (see Figure 1). Opposing tubes must be filled to the same level with liquid of the same density.

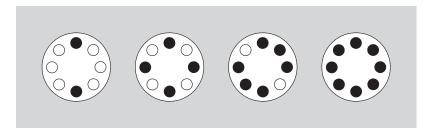
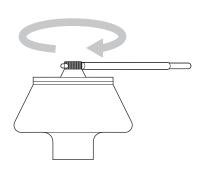


Figure 1. Arranging Tubes in the Rotor. Two, four, six, or eight tubes can be centrifuged per run if they are arranged in the rotor as shown.

3. Complete loading by placing the correct floating spacers over the tubes.

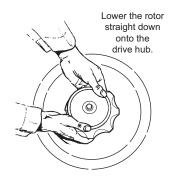


IIII NOTE

Place filled tubes in at least two opposing cavities. Make sure that cavities in use also have the proper spacers inserted before installing the rotor lid. Do not put spacers in cavities that do not contain tubes.

4. Put the lid in place and tighten, as firmly as possible, with the handle tool (356959) provided. Remove the handle tool.

OPERATION



- 1. Carefully place the rotor on the drive hub.
- 2. Refer to the instrument instruction manual for ultracentrifuge operation.
- 3. For additional operating information, see the following:
 - RUN TIMES, page 9, for using k factors to adjust run durations
 - RUN SPEEDS, page 10, for information about speed limitations

- SLOW ACCELERATION/DECELERATION, page 12, for information about using slow acceleration and deceleration for gradient stability when preformed gradients are used
- SELECTING CsCl GRADIENTS, page 13, for methods to avoid CsCl precipitation during centrifugation

REMOVAL AND SAMPLE RECOVERY



CALITION

If disassembly reveals evidence of leakage, you should assume that some fluid escaped the rotor. Apply appropriate decontamination procedures to the centrifuge and accessories.

- 1. Remove the rotor from the centrifuge by lifting it straight up and off the drive hub.
- 2. Remove the rotor lid.
- 3. Use the removal tool (361668) to remove the spacers and tubes.



The extremely high centrifugal forces of an ultra-high-speed rotor cause a slight vacuum to build up in the tube cavities, occasionally resulting in a suction effect when removing the tubes from the rotor. Some tube deformation may also occur. A brief delay (approximately 5 minutes) after the rotor comes to rest before removing the tubes will make tube removal easier. If you experience difficulty in removing the tubes from the rotor, use a gentle twisting or rocking motion, and remove the tube slowly to avoid sample mixing.



Removal Tool (361668)

TUBES

The Type 100 Ti rotor uses only the Quick-Seal tubes and accessories listed in Table 1; be sure to use only those items. Refer to *Rotors and Tubes* for information on the chemical resistances of tube and accessory materials.

Table 1. Available Tubes for the Type 100 Ti Rotor. Use only the items listed here and observe maximum fill volumes and speeds shown.

Tube		Required Accessory		Max.	
Dimensions and Volume	Description	Part Number	Description	Part Number	Speed/ RCF/ <i>k</i> Factor
13 × 65 mm 6.7 mL	Quick Seal polyallomer, bell-top	363650 (pkg/50)	Noryl ^a floating spacer	393636 (pkg/8)	100 000 rpm 802 000 × <i>g</i> 17
13 × 57 mm 6.0 mL	Quick-Seal polyallomer, bell-top	344619 (pkg/50)	Noryl floating spacer	365470 (pkg/8)	100 000 rpm 802 000 × <i>g</i> 15
13 × 51 mm 5.1 mL	Quick-Seal polyallomer, bell-top	362248 ^b (pkg/50)	Noryl floating spacer	360270 (pkg/8)	100 000 rpm 802 000 × <i>g</i> 13.5
13 × 30 mm 3.5 mL	Quick-Seal polyallomer, bell-top	349621° (pkg/50)	Noryl floating spacer	360270 (pkg/8)	100 000 rpm 802 000 × <i>g</i> 9.4
13 × 25 mm 2.0 mL	Quick-Seal polyallomer, bell-top	345829 ^d (pkg/50)	Noryl floating spacer	360270 (pkg/8)	100 000 rpm 802 000 × <i>g</i> 7

^a Noryl is a registered trademark of GE Plastics.

^b Tube 362248 is also available as part of *g*-Max Kit 363960.

^c Tube 349621 is also available as part of *g*-Max Kit 363961.

^d Tube 345829 is also available as part of *g*-Max Kit 363962.



Temperature Limits

- Plastic tubes have been centrifuge tested for use at temperatures between 2 and 25°C. For centrifugation at other temperatures, pretest tubes under anticipated run conditions.
- If tubes are frozen before use, make sure that they are thawed to at least 2°C prior to centrifugation.



Quick-Seal® Tubes

Quick-Seal tubes must be sealed prior to centrifugation. These tubes are heat sealed and do not need caps; however, spacers are required on top of the tubes when they are loaded into the rotor.

- Fill Quick-Seal tubes leaving a *small* bubble of air at the base of the neck. Do not leave a large air space—too much air can cause excessive tube deformation.
- Refer to *Rotors and Tubes* for detailed information on the use and care of Quick-Seal tubes.

Some of the tubes listed in Table 1 are part of the g-MaxTM system. The g-Max system uses a combination of small bell-top Quick-Seal tubes and floating spacers (also called g-Max spacers). This means that you can run the shorter tubes listed in the table in the Type 100 Ti rotor without reduction in g force. Additional information about the g-Max system is available in publication DS-709.

RUN TIMES

TIME HR:MIN

00:30

The k factor of the rotor is a measure of the rotor's pelleting efficiency. (Beckman Coulter has calculated the k factors for all of its preparative rotors at maximum rated speed and using full tubes.) The k factor is calculated from the formula:

$$k = \frac{\ln(r_{\text{max}}/r_{\text{min}})}{\omega^2} \times \frac{10^{13}}{3600}$$
 (1)

where ω is the angular velocity of the rotor in radians per second ($\omega = 0.105 \times \text{rpm}$), r_{max} is the maximum radius, and r_{min} is the minimum radius.

After substitution:

$$k = \frac{(2.533 \times 10^{11}) \ln(r_{\text{max}}/r_{\text{min}})}{\text{rpm}^2}$$
 (2)

Use the k factor in the following equation to estimate the run time t (in hours) required to pellet particles of known sedimentation coefficient s (in Svedberg units, S).

$$t = \frac{k}{s} \tag{3}$$

Run times can be estimated for centrifugation at less than maximum speed by adjusting the k factor as follows:

$$k_{\text{adj}} = k \left(\frac{100\ 000}{\text{actual run speed}} \right)^2 \tag{4}$$

Run times can also be estimated from data established in prior experiments if the k factor of the previous rotor is known. For any two rotors, a and b:

$$\frac{t_{\rm a}}{t_{\rm b}} = \frac{k_{\rm a}}{k_{\rm b}} \tag{5}$$

For more information on *k* factors see *Use of* k *Factor for Estimating Run Times from Previously Established Run Conditions* (publication DS-719).

RUN SPEEDS

SPEED RPM/RCF

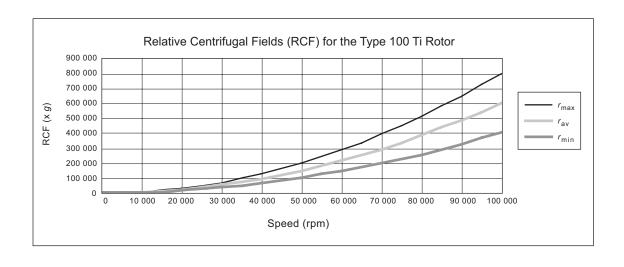
100 000 RPM

The centrifugal force at a given radius in a rotor is a function of speed. Comparisons of forces between different rotors are made by comparing the rotors' relative centrifugal fields (RCF). When rotational speed is adjusted so that identical samples are subjected to the same RCF in two different rotors, the samples are subjected to the same force. The RCF at a number of rotor speeds is provided in Table 2.

Table 2. Relative Centrifugal Fields for the Type 100 Ti Rotor. Entries in this table are calculated from the formula $RCF = 1.12r (RPM/1000)^2$ and then rounded to three significant digits.

5.	Relat			
Rotor Speed (rpm)	At r _{max} (71.6 mm)	At <i>r</i> _{av} (55.5 mm)	At r _{min} (39.5 mm)	k Factor*
100 000	802 000	622 000	442 000	15
95 000	724 000	561 000	399 000	17
90 000	650 000	504 000	358 000	19
85 000	580 000	449 000	320 000	21
80 000	513 000	398 000	283 000	24
75 000	451 000	350 000	249 000	27
70 000	393 000	305 000	217 000	31
65 000	339 000	263 000	187 000	36
60 000	289 000	224 000	159 000	42
55 000	243 000	188 000	134 000	50
50 000	201 000	155 000	111 000	60
45 000	162 000	126 000	89 600	74
40 000	128 000	99 500	70 800	94
35 000	98 200	76 200	54 200	123
30 000	72 200	55 900	39 800	167
25 000	50 100	38 900	27 700	241

^{*}Calculated for all Beckman Coulter preparative rotors as a measure of the rotor's relative efficiency in pelleting sample in water at 20° C.



Speeds must be reduced under the following circumstances:

1. If nonprecipitating solutions more dense than 1.2 g/mL are centrifuged, the maximum allowable run speed must be reduced according to the following equation:

reduced maximum speed = (100 000 rpm)
$$\sqrt{\frac{1.2 \text{ g/mL}}{\rho}}$$
 (6)

where ρ is the density of the tube contents. This speed reduction will protect the rotor from excessive stresses due to the added tube load. *Note, however, that the use of this formula may still produce maximum speed figures that are higher than the limitations imposed by the use of certain tubes or adapters.* In such cases, use the lower of the two figures.

2. Further speed limits must be imposed when CsCl or other self-forming-gradient salts are centrifuged, as equation (6) does not predict concentration limits/speeds that are required to avoid precipitation of salt crystals. Precipitation during centrifugation would alter the density distribution of CsCl and this would change the position of the sample bands. Figures 2 and 3, together with the description and examples below, show how to reduce run speeds when using CsCl gradients.

SLOW ACCELERATION/DECELERATION

Vertical banding of sample and gradient formation occurs with centrifugation. With deceleration, tube contents reorient back to horizontal position. For gradient stability *when preformed gradients are used*, select slow acceleration as follows:

ACCEL PROFILES

- in Optima L-XP and XL series ultracentrifuges, select a slow acceleration profile.
- in Optima L series ultracentrifuges, select SLOW ACCEL.

For the stability of *all gradients* during deceleration, do the following:

DECEL PROFILES

- in Optima L-XP and XL series ultracentrifuges, select a slow deceleration profile.
- in Optima L series ultracentrifuges, select SLOW DECEL.

SELECTING CsCl GRADIENTS



Rotor speed is used to control the slope of a CsCl density gradient, and must be limited so that CsCl precipitation is avoided. Precipitation during centrifugation would alter density distribution, and this would change the position of the sample bands. Curves in Figures 2 and 3 are provided up to the maximum rated speed of the rotor.

IIII NOTE

The curves in Figures 2 and 3 are for solutions of CsCl salt dissolved in distilled water only. If other salts are present in significant concentrations, the overall CsCl concentration may need to be reduced.

Speed and density combinations that intersect on or below the curves in Figure 3 ensure that CsCl will not precipitate during centrifugation in the Type 100 Ti rotor. Curves are provided at two temperatures: 20°C (black curves) and 4°C (gray curves).

The reference curves in Figure 3 show gradient distribution at equilibrium. Each curve in Figure 3 is within the density limits allowed for the Type 100 Ti rotor: each curve was generated for a single run speed using the maximum allowable homogeneous CsCl densities (one for each fill level) that avoid precipitation at that speed. (The gradients in Figure 3 can be generated from step or linear gradients, or from homogeneous solutions. But the total amount of CsCl in solution must be equivalent to a homogeneous solution corresponding to the concentrations specified in Figure 2.) Figure 3 can also be used to approximate the banding positions of sample particles.

ADJUSTING FILL VOLUMES

Figures 2 and 3 show that several fill volumes are possible in a tube. (The curves shown are for the 6-mL tube, 344619. The 3.5-mL tube, 349621, is half as long as the tube shown; therefore, a *full* 3.5-mL tube would be approximately the same as a *half-full* 6.0-mL tube.) If a tube is partially filled with gradient solution, float mineral oil (or some other low-density, immiscible liquid) on top of the tube contents to fill the tube to its maximum volume. Note that for a given CsCl density, as the fill level decreases the maximum allowable speed increases. Partial filling may be desirable when there is little sample or when you wish to shorten the run time.

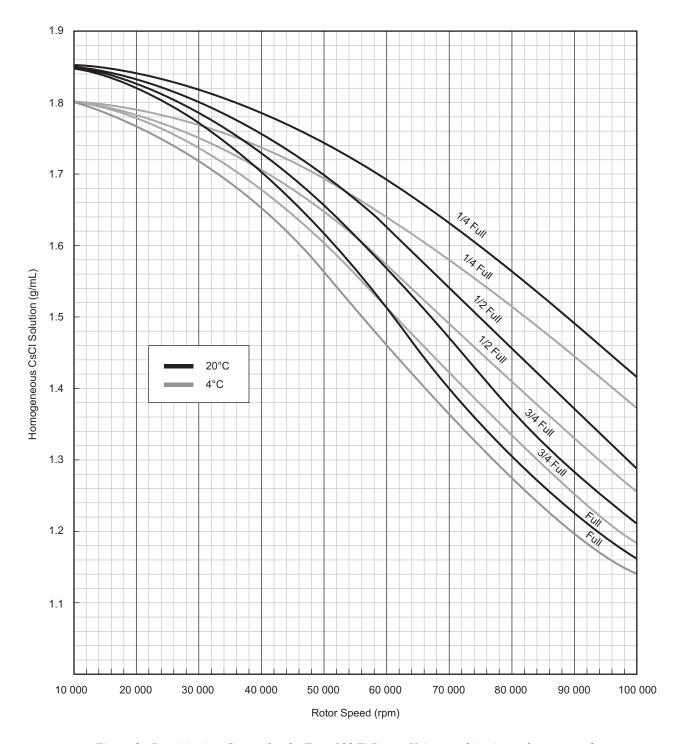


Figure 2. Precipitation Curves for the Type 100 Ti Rotor. Using combinations of rotor speeds and homogeneous CsCl solution densities that intersect on or below these curves ensures that CsCl will not precipitate during centrifugation.

The curves shown here and in Figure 3 represent the 6.0-mL Quick-Seal tube.

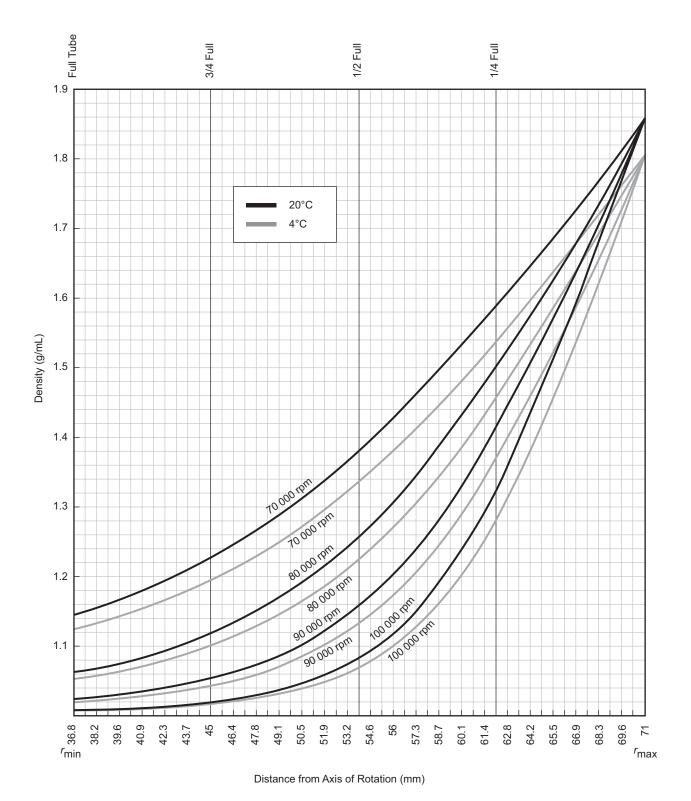
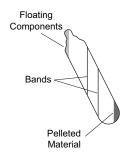
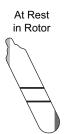


Figure 3. CsCl Gradients at Equilibrium for the Type 100 Ti Rotor. Centrifugation of homogeneous CsCl solutions at the maximum allowable speeds (from Figure 2) results in gradients presented here. For example, a *half-filled* tube of 1.40-g/mL homogeneous CsCl solution at 20°C may be centrifuged at 88 000 rpm (see Figure 2). The same solution in a *three-quarter-filled* tube may be centrifuged no faster than 78 000 rpm. A tube full of the 1.40-g/mL CsCl solution may be centrifuged no faster than 70 000 rpm.

TYPICAL EXAMPLES FOR DETERMINING CsCI RUN PARAMETERS

At Speed







Example A: Knowing homogeneous CsCl solution density (such as 1.50 g/mL) and approximate particle buoyant densities (1.60 and 1.70 g/mL), at 4°C, where will particles band?

- 1. In Figure 2, find the curve that corresponds to the desired run temperature (4°C). The maximum allowable rotor speed is determined from the point where this curve intersects the homogeneous CsCl density (68 000 rpm).
- 2. In Figure 3, sketch in a horizontal line corresponding to each particle's buoyant density.
- 3. Mark the point in the figure where each particle density intersects the curve corresponding to the selected run speed and temperature. (Gradient curves not shown in Figure 3 can be interpolated.)
- 4. Particles will band at these locations across the tube diameter at equilibrium during centrifugation.

In this example, particles will band about 66.3 and 64.2 mm from the axis of rotation, about 2.1 mm of centerband-to-centerband separation at the rotor's 26-degree tube angle. When the tube is removed from the rotor and held upright (vertical and stationary), there will be about 2.3 mm of centerband-to-centerband separation. This interband distance, $d_{\rm up}$, can be calculated from the formula:

$$d_{\rm up} = \frac{d_{\theta}}{\cos \theta} \tag{7}$$

where $d\theta$ is the interband distance when the tube is held at an angle, θ , in the rotor.

Example B: Knowing particle buoyant densities (e.g., 1.65 and 1.70 g/mL), how do you achieve good separation?

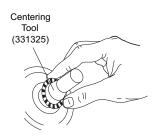
1. In Figure 3, sketch in a horizontal line corresponding to each particle's buoyant density.

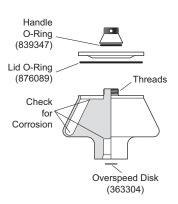
- 2. Select the curve at the desired temperature (20°C) that gives the best particle separation.
- 3. Note the run speed along the selected curve (70 000 rpm).
- 4. From Figure 2, select the maximum homogeneous CsCl density (in this case, 1.41 g/mL) that corresponds to the temperature and run speed established above. These parameters will provide the particle-banding pattern selected in Step 2.

In this example, particles will band at about 64 and 65.2 mm from the axis of rotation (about 1.2 mm apart). When the tube is held upright there will be about 1.3 mm of center-of-band to center-of-band separation.

CARE AND MAINTENANCE

MAINTENANCE







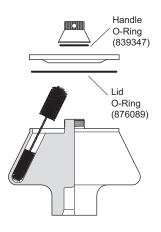
Do not use sharp tools on the rotor that could cause scratches in the rotor surface. Corrosion begins in scratches and may open fissures in the rotor with continued use.

- Regularly inspect the overspeed disk. If it is scratched, damaged, or missing, replace it. Replacement instructions are in *Rotors and Tubes*.
- Regularly lubricate the metal threads in the rotor and handle with a thin, even coat of Spinkote lubricant. Failure to keep these threads lubricated can result in damaged threads.
- Regularly apply silicone vacuum grease to the O-rings. Replace O-rings about twice a year or whenever worn or damaged.

Refer to Appendix A in *Rotors and Tubes* for the chemical resistances of rotor and accessory materials. Your Beckman Coulter representative provides contact with the Field Rotor Inspection Program and the rotor repair center.

CLEANING





Wash the rotor and rotor components immediately if salts or other corrosive materials are used or if spillage has occurred. Do not allow corrosive materials to dry on the rotor.

Under normal use, wash the rotor frequently (at least weekly) to prevent buildup of residues.

- 1. Remove the O-rings before washing.
- 2. Wash the rotor and lid in a mild detergent, such as Beckman Solution 555TM, that won't damage the rotor. The Rotor Cleaning Kit contains two plastic-coated brushes and two quarts of Solution 555 (339555) for use with rotors and accessories. Dilute the detergent 10 to 1 with water.



Do not wash rotor components in a dishwasher. Do not soak in detergent solution for long periods, such as overnight.

- 3. Rinse the cleaned rotor and components with distilled water.
- 4. Air-dry the rotor and lid upside down. *Do not use acetone to dry the rotor*.
- 5. Apply a thin, even coat of silicone vacuum grease (335148) to the lid O-ring, then replace it in the groove in the outer rim of the lid.

Clean metal threads every 6 months, or as necessary. Use a brush and concentrated Solution 555. Rinse and dry thoroughly, then lubricate lightly but evenly with Spinkote to coat all threads.

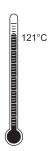
Periodically remove the O-rings and wipe clean as necessary. Clean the O-ring grooves with a cotton-tipped swab. Reapply a light film of silicone vacuum grease.

DECONTAMINATION



If the rotor or other components are contaminated with toxic or pathogenic materials, follow appropriate decontamination procedures as outlined by your laboratory safety officer. Check Appendix A in *Rotors and Tubes* to be sure the decontamination method will not damage any part of the rotor.

STERILIZATION AND DISINFECTION



- The rotor and all rotor components can be autoclaved at 121°C for up to an hour. Remove the lid from the rotor and place the rotor, lid, and spacers in the autoclave upside down.
- Ethanol (70%)* or hydrogen peroxide (6%) may be used on all rotor components, including those made of plastic. Bleach (sodium hypochlorite) may be used, but may cause discoloration of anodized surfaces. Use the minimum immersion time for each solution, per laboratory standards.

While Beckman Coulter has tested these methods and found that they do not damage the rotor or components, no guarantee of sterility or disinfection is expressed or implied. When sterilization or disinfection is a concern, consult your laboratory safety officer regarding proper methods to use.

Refer to publication IN-192 for tube sterilization procedures. *Quick-Seal tubes are disposable and should be discarded after a single use.*

STORAGE

When it is not in use, store the rotor in a dry environment (not in the instrument) with the lid removed to allow air circulation so moisture will not collect in the tube cavities.

^{*} Flammability hazard. Do not use in or near operating ultracentrifuges.

RETURNING A ROTOR



Before returning a rotor or accessory for any reason, prior permission (a Returned Goods Authorization form) must be obtained from Beckman Coulter, Inc. This RGA form may be obtained from your local Beckman Coulter sales office, and should contain the following information:

- serial number,
- history of use (approximate frequency of use),
- reason for the return,
- original purchase order number, billing number, and shipping number, if possible,
- name and phone number of the person to be notified upon receipt of the rotor or accessory at the factory,
- name and phone number of the person to be notified about repair costs, etc.

To protect our personnel, it is the customer's responsibility to ensure that all parts are free from pathogens and/or radioactivity. Sterilization and decontamination must be done before returning the parts. Smaller items (such as tubes, bottles, etc.) should be enclosed in a sealed plastic bag.

All parts must be accompanied by a note, plainly visible on the outside of the box or bag, stating that they are safe to handle and that they are not contaminated with pathogens or radioactivity. Failure to attach this notification will result in return or disposal of the items without review of the reported problem.

Use the address label printed on the RGA form when mailing the rotor and/or accessories.

Customers located outside the United States should contact their local Beckman Coulter office.

SUPPLY LIST

Contact Beckman Coulter Sales (1-800-742-2345 in the United States; worldwide offices are listed on the back cover of this manual) or see the Beckman Coulter *Ultracentrifuge Rotors, Tubes & Accessories* catalog (BR-8101) for detailed information on ordering parts and supplies. For your convenience, a partial list is given below.



Publications referenced in this manual can be obtained by calling Beckman Coulter at 1-800-742-2345 in the United States, or by contacting your local Beckman Coulter office.

REPLACEMENT ROTOR PARTS

Type 100 Ti rotor assembly	363013
Rotor handle	363039
O-ring (lid)	876089
O-ring (handle)	839347
Overspeed disk (100 000 rpm)	363304

OTHER

Tubes and spacers see Table 1
Quick-Seal Cordless Tube Topper kit, 60 Hz
Quick-Seal Cordless Tube Topper kit, 50 Hz (Europe) 358313
Quick-Seal Cordless Tube Topper kit, 50 Hz (Great Britain) 358314
Quick-Seal Cordless Tube Topper kit, 50 Hz (Australia)
Quick-Seal Cordless Tube Topper kit, 50 Hz (Canada)
Tube Topper rack (16-mm dia. tubes)
Handle tool
Quick-Seal tube removal tool
Spinkote lubricant (1 oz)
Silicone vacuum grease (2 oz)
Rotor Cleaning Kit
Beckman Solution 555 (1 qt)
Rotor cleaning brush
Centering tool (for replacing overspeed disk)

ULTRACENTRIFUGE ROTOR WARRANTY

All Beckman Coulter ultracentrifuge Fixed Angle, Vertical Tube, Near Vertical Tube, Swinging Bucket, and Airfuge rotors are warranted against defects in materials or workmanship for the time periods indicated below, subject to the Warranty Conditions stated below.

Preparative Ultracentrifuge Rotors 5 years — No Proration

Analytical Ultracentrifuge Rotors . . . 5 years — No Proration

ML and TL Series Ultracentrifuge
Rotors 5 years — No Proration

Airfuge Ultracentrifuge Rotors . . . 1 year — No Proration

For Zonal, Continuous Flow, Component Test, and Rock Core ultracentrifuge rotors, see separate warranty.

Warranty Conditions (as applicable)

- This warranty is valid for the time periods indicated above from the date of shipment to the original Buyer by Beckman Coulter or an authorized Beckman Coulter representative.
- This warranty extends only to the original Buyer and may not be assigned or extended to a third person without written consent of Beckman Coulter.
- 3) This warranty covers the Beckman Coulter Centrifuge Systems only (including but not limited to the centrifuge, rotor, and accessories) and Beckman Coulter shall not be liable for damage to or loss of the user's sample, non-Beckman Coulter tubes, adapters, or other rotor contents.
- 4) This warranty is void if the Beckman Coulter Centrifuge System is determined by Beckman Coulter to have been operated or maintained in a manner contrary to the instructions in the operator's manual(s) for the Beckman Coulter Centrifuge System components in use. This includes but is not limited to operator misuse, abuse, or negligence regarding indicated maintenance procedures, centrifuge and rotor classification requirements, proper speed reduction for the high density of certain fluids, tubes, and tube caps, speed reduction for precipitating gradient materials, and speed reduction for high-temperature operation.
- 5) Rotor bucket sets purchased concurrently with or subsequent to the purchase of a Swinging Bucket Rotor are warranted only for a term co-extensive with that of the rotor for which the bucket sets are purchased.
- 6) This warranty does not cover the failure of a Beckman Coulter rotor in a centrifuge not of Beckman Coulter manufacture, or if the rotor is used in a Beckman Coulter centrifuge that has been modified without the written permission of Beckman Coulter, or is used with carriers, buckets, belts, or other devices not of Beckman Coulter manufacture.
- 7) Rotor parts subject to wear, including but not limited to rotor O-rings, VTi, NVTTM, TLV, MLN, and TLN rotor tube cavity plugs and gaskets, tubing, tools, optical overspeed disks, bearings, seals, and lubrication are excluded from this warranty and should be frequently inspected and replaced if they become worn or damaged.
- Keeping a rotor log is not mandatory, but may be desirable for maintenance of good laboratory practices.

Repair and Replacement Policies

- If a Beckman Coulter rotor is determined by Beckman Coulter to be defective, Beckman Coulter will repair or replace it, subject to the Warranty Conditions. A replacement rotor will be warranted for the time remaining on the original rotor's warranty.
- 2) If a Beckman Coulter centrifuge is damaged due to a failure of a rotor covered by this warranty, Beckman Coulter will supply free of charge (i) all centrifuge parts required for repair (except the drive unit, which will be replaced at the then current price less a credit determined by the total number of revolutions or years completed, provided that such a unit was manufactured or rebuilt by Beckman Coulter), and (ii) if the centrifuge is currently covered by a Beckman Coulter warranty or Full Service Agreement, all labor necessary for repair of the centrifuge.
- 3) If a Beckman Coulter rotor covered by this warranty is damaged due to a malfunction of a Beckman Coulter ultracentrifuge covered by an Ultracentrifuge System Service Agreement, Beckman Coulter will repair or replace the rotor free of charge.
- 4) If a Beckman Coulter rotor covered by this warranty is damaged due to a failure of a Beckman Coulter tube, bottle, tube cap, spacer, or adapter, covered under the Conditions of this Warranty, Beckman Coulter will repair or replace the rotor and repair the instrument as per the conditions in policy point (2) above, and the replacement policy.
- 5) Damage to a Beckman Coulter rotor or instrument due to the failure or malfunction of a non-Beckman Coulter tube, bottle, tube cap, spacer, or adapter is not covered under this warranty, although Beckman Coulter will assist in seeking compensation under the manufacturer's warranty.

Disclaimer

IT IS EXPRESSLY AGREED THAT THE ABOVE WARRANTY SHALL BE IN LIEU OF ALL WARRANTIES OF FITNESS AND OF THE WARRANTY OF MERCHANTABILITY AND BECKMAN COULTER, INC. SHALL HAVE NO LIABILITY FOR SPECIAL OR CONSEQUENTIAL DAMAGES OF ANY KIND WHATSOEVER ARISING OUT OF THE MANUFACTURE, USE, SALE, HANDLING, REPAIR, MAINTENANCE, OR REPLACEMENT OF THE PRODUCT.

Factory Rotor Inspection Service

Beckman Coulter, Inc., will provide free mechanical and metallurgical inspection in Palo Alto, California, USA, of any Beckman Coulter rotor at the request of the user. (Shipping charges to Beckman Coulter are the responsibility of the user.) Rotors will be inspected in the user's laboratory if the centrifuge in which they are used is covered by an appropriate Beckman Coulter Service Agreement. Contact your local Beckman Coulter office for details of service coverage or cost.

Before shipping, contact the nearest Beckman Coulter Sales and Service office and request a Returned Goods Authorization (RGA) form and packaging instructions. Please include the complete rotor assembly, with buckets, lid, handle, tube cavity caps, etc. A SIGNED STATEMENT THAT THE ROTOR AND ACCESSORIES ARE NON-RADIOACTIVE, NON-PATHOGENIC, NON-TOXIC, AND OTHERWISE SAFE TO SHIP AND HANDLE IS REQUIRED.

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