

# Biallelic CRISPR-Cas9 editing of gene associated with coat color in microinjected bovine zygotes reaching the blastocyst stage

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## Background

CRISPR studies in cattle have focused on cell editing and subsequent cloning (SCNT), attempting founder animal production. Further studies in cattle have been elucidating the role of cell differentiation genes in zygote using both micro-injection and SCNT [1,2]. Editing of these genes led to developmental arrest before reaching blastocyst stage as well as lower blastocyst rate. Additional efforts have been performed to reduce mosaicism by changing the injection time post-fertilization.

COPA is involved in ER-Golgi transport and its mutation is associated with coat color variation in cattle [3], but also in lung diseases and arthritis in humans [4] and increased proliferation, migration and invasion of cancer cells *in vitro* [5].

## Aim

We investigated the feasibility of CRISPR-Cas9 gene editing via micro-injection and electroporation in a conventional IVF protocol to derive edited blastocysts.

## Methods

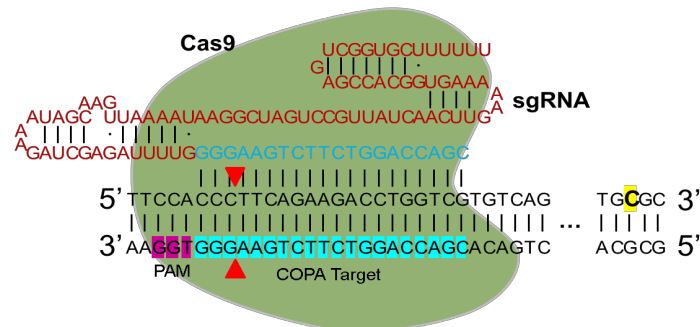
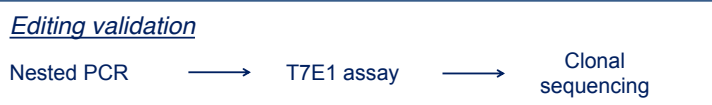
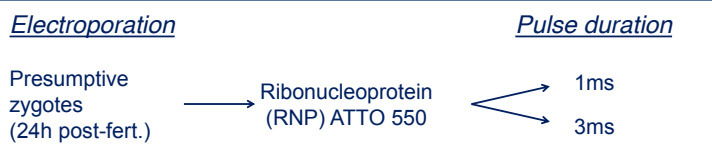
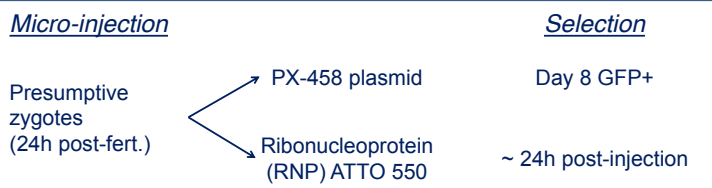


Figure 1 – Schematic representation of the COPA sgRNA target. The missense mutation associated with coat color in cattle is highlighted in yellow.



## References

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- Simmet K, Zakharichenko V, Philippo-Masser J, Blum H, Klymiuk N, Wolf E. *OCT4/POU5F1* is required for *NANOG* expression in bovine blastocysts. PNAS 115(11) 2018.
- Dorshorst B, Henegar C, Liao X, Almen M S, Rubin CJ, Ito S, Wakamatsu K, Stothard P, Van Doormaaf B, Plastow G, Barsh G S, Andersson L. Dominant red coat color in holstein cattle is associated with a missense mutation in the coatomer protein complex, subunit alpha (COPA) gene. PLOS ONE 10(6) 2015.
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## Results

### Fluorescence is not a good editing selection tool

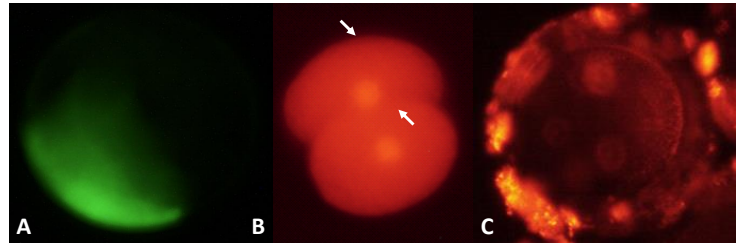


Figure 2 – Fluorescence selection of CRISPR injected presumptive zygotes (A, B) as well as electroporated zygotes (C). Arrows indicate the localization of the RNP effector complex in the nucleus.

### Electroporation of RNP yields the highest editing rate

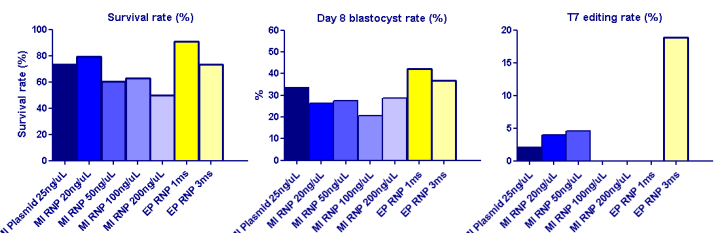


Figure 3 – Survival, blastocyst and T7 editing rate of both micro-injected (MI) and electroporated (EP) groups at various concentrations as well as pulse length.

### COPA biallelic edited zygotes can reach blastocyst stage

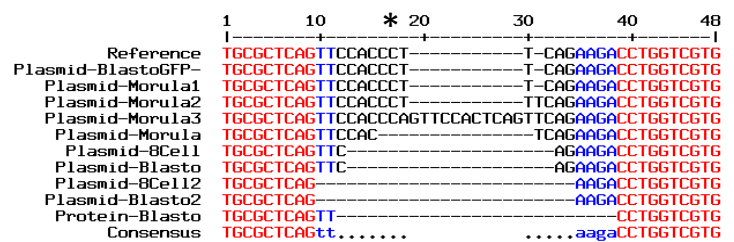


Figure 4 – Characterization of editing events in plasmid and protein injected embryos. \* represents the sgRNA guided restriction site.

## Conclusion

Microinjection and electroporation of CRISPR plasmids and protein in presumptive zygotes is possible and mosaic editing as well as biallelic editing can occur from this procedure. Even if survival rates and developmental rates are similar, electroporation of RNP seems to yield a higher amount of editing in blastocysts.

Since coatomer protein complex subunit alpha (COPA) acts as a protein carrier, it is possible that homozygous knock-out causes developmental arrest in embryos. Homology-directed repair should ensure proper missense mutation of the gene in future founder animals.